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SUGAR MODIFIED OLIGONUCLEOTIDES

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Abstract. The synthesis and enzymatic stability of oligonucleotides with $1-(2,3-dideoxy-\beta-D-\underline{erythro}-hexopyranosyl)thymine and <math>(\underline{S})-9-(3,4-dihydroxybutyl)$ adenine as monomeric units are described.

In view of the recent interest in antisense oligonucleotides as antiviral or antitumor agents, we became interested in the synthesis of nuclease-resistant oligonucleotides. Because modifications of the phosphate moiety have been the subject of intensive research (e.g. the synthesis of phosphorothioates, phosphorodithioates and methylphosphonates), we concentrated our efforts on sugar-modified oligonucleotides. We describe here the synthesis and enzymatic stability of oligonucleotides containing $1-(2,3-dideoxy-\beta-D-erythro-hexopyranosyl)$ thymine 1 and $(\underline{S})-9-(3,4-dihydroxybutyl)$ adenine 6 as monomers.

The condensation of 3,4,6-tri- \underline{O} -acetyl-D-glucal and bistrimethyl-silylthymine in the presence of trimethylsilyl trifluoromethanesulfonate yielded 1-(4,6-di- \underline{O} -acetyl-2,3-dideoxy- β -D- \underline{e} -rythro-hex-2-enopyranosyl)thymine. This gave compound 1 following hydrogenation and deacetylation. The synthesis of the acyclic monomer started from (\underline{S})-1,2- \underline{O} -isopropylidene-1,2,4-butanetriol. After mesylation we obtained (\underline{S})-4- \underline{O} -methanesulfonyl-1,2- \underline{O} -isopropylidene-1,2,4-butanetriol. The reaction of this product with adenine in the presence of sodium hydride afforded (\underline{S})-9-(3,4- \underline{O} -isopropylidene-3,4-dihydroxybutyl)adenine. Subsequent protection of the exocyclic aminofunction of adenine with a benzoylgroup followed by acidic hydrolysis yielded compound 7.

The primary hydroxyl group of both 1 and 7 was protected with a monomethoxytrityl group to afford 2 and 8. The secondary hydroxyl group of 2 and 8 was functionalised to yield the phosphoramidite 9 and the H-

phosphonates 3 and 10. These were used as building blocks to synthesize several dimers using the phosphoramidite or the H-phosphonate approach. The following dimers were synthesized: T*pT*; T*pT; TpT*; A*pA*; A*pA; $ApA^* [T^* = 1, A^* = 6, T = thymidine, A = 2'-deoxyadenosine]. Compared$ with the dimers TpT and ApA, all dimer analogues have an improved stability against the hydrolytic activity of nuclease S_1 , snake venom phosphodiesterase and bovine spleen phosphodiesterase. For the synthesis of longer oligonucleotides we made use of the analogs 4 and 11 functionalized with a dimethoxytrityl protecting group. We also prepared 5 and 12 which were linked to a LCAA-CPG support. Synthesis was done on an Applied Biosystems 381A model using the H-phosphonate approach. The stepwise coupling yields were around 95% for the unmodified nucleotides and around 90% for the modified nucleotides. Following oligonucleotides were prepared: $T^*T_{11}T^*$, $T^*T^*T_0T^*T^*$, $T^*T_5T^*T_5T^*$, $T_6T^*T_6$, T^*_{13} and the adenosine oligomers $A^*A_{11}A^*$, $A^*A^*A_9A^*A^*$, $A^*A_5A^*A_5A^*$, $A_6A^*A_6$, A^*_{13} . Determination of thermal stabilities of these oligomers with the natural A_{13} and T_{13} are underway.

- 1 R₁= R₂= H
 2 R₁= MMTr, R₂= H
 7 R₁= R₂= H, R₃= C-C₆H₅
 3 R₁= MMTr, R₂= P-O-Et₃NH+
 H
 8 R₁= MMTr, R₂= H, R₃= C-C₆H₅
 O
 4 R₁= DMTr, R₂= P-O-Et₃NH+
 H
 9 R₁= MMTr, R₂= PO-Et₃NH+
 COCH₂CH₂CN
 O
 10 R₁= MMTr, R₂= P-O-Et₃NH+, R₃= C-C₆H₅
 O
 10 R₁= MMTr, R₂= P-O-Et₃NH+, R₃= C-C₆H₅
 O
 10 R₁= MMTr, R₂= P-O-Et₃NH+, R₃= C-C₆H₅
 O
 10 R₁= MMTr, R₂= P-O-Et₃NH+, R₃= C-C₆H₅

 - 11 $R_1 = DMTr, R_2 = \overset{H}{P} O^*Et_3NH^+, R_3 = \overset{C}{C} \overset{C}{C}_6H_5$
 - 12 $R_1 = DMTr$, $R_2 = CCH_2CH_2COOH$, $R_3 = CCGH_5$